

# COVID-19 Next Generation Sequencing Specimen Collection and Submission Instructions

# Coordination with Epidemiologists

Providers interested in submitting samples for SARS-CoV-2 next generation sequencing (NGS) must coordinate with their Local and/or Regional Public Health Department (<a href="https://www.dshs.texas.gov/regions/2019-nCoV-Local-Health-Entities/">https://www.dshs.texas.gov/regions/2019-nCoV-Local-Health-Entities/</a>). Local and Regional Public Health Departments must seek sequencing approval from Texas DSHS EAIDU epidemiologists (<a href="mailto:EAIDU-Coronavirus@dshs.texas.gov">EAIDU-Coronavirus@dshs.texas.gov</a>) to initiate the submission process.

### Criteria for Prioritized Specimens

- a. Specimens from patients who already have a positive COVID-19 PCR lab result; a PCR positive sample must be available for sequencing
- b. Priority specimens for <u>reinfection evaluation</u> include the following:
  - Specimens from symptomatic or asymptomatic COVID-19 patients who have 2
    positive PCR tests separated by at least 90 days and stored clinical samples or RNA
    extracts are available for both tests
- c. Priority specimens for genetic variant surveillance include the following:
  - Symptomatic or asymptomatic COVID-19 patients who are close contacts with a case of known infection with a SARS-CoV-2 genetic variant of concern
  - Patients with a history of international travel to a location where there is known, or suspected, community spread of one or more SARS-CoV-2 genetic variants of concern
  - Patients with an extremely severe or unusual COVID-19 illness presentation
  - Patients who are fully vaccinated with an FDA authorized COVID-19 vaccine, with the
    most recent dose occurring at least 2 weeks prior to illness onset and no SARS-CoV2 RNA or antigen detected on a respiratory specimen collected <45 days before the
    most recent positive test.</li>
  - Patients who developed COVID-19 in congregate living settings with attack rates over 75% within a 2-week period, where up to 5 samples will be sequenced per outbreak
  - Other outbreak specimens where there is reason to suspect an infection with a SARS-CoV-2 genetic variant of concern.
  - Specimens from COVID-19 associated pediatric deaths
  - Specimens from potential reinfections with at least 2 positive PCR results that meet all other sequencing criteria (this includes samples where initial positive PCR test sample is unavailable)

## Specimen Types

- All specimens submitted for Next Generation Sequencing (NGS) must have been previously tested by PCR methods and confirmed to be positive for COVID-19
- The PCR results should have target gene PCR CT value of ≤ 30
- The ONLY samples that can be sequenced with CT values > 30 are vaccine breakthrough and reinfection cases

#### RNA Extracts (preferred)

• minimum volume required: 30 µl

### Clinical Specimens

- Nasopharyngeal (NP) swabs in viral transport media (VTM)
- Oropharyngeal (OP) swabs and Nasal swabs are also acceptable
- Specimen collected in Universal Transport Media (UTM), Amies, or 0.85% Saline are also acceptable
- Minimum volume required: 0.8 mL (volume as low as 0.4mL may be accepted for testing but may not be sufficient for completion of all tests required)



# **COVID-19 Next Generation Sequencing Specimen Collection and Submission Instructions**

Specimen Collection, Handling, and Shipping	<ul> <li>RNA extracts and clinical specimens should be stored frozen at -70°C and must be shipped frozen on dry ice for overnight delivery.</li> <li>Complete the line list. All fields marked with an asterisk (*) must be filled. Ship the specimens with a hard copy of the completed line list to the DSHS Laboratory.</li> <li>If sending both RNA extracts and clinical specimens in the same shipment, enter all RNA extract specimen information at the top of the line list followed by clinical specimen information.</li> <li>Label the tubes with the specimen ID and date of collection. The identifiers on the tubes must match those entered on the line list.</li> <li>If sending RNA extracts and clinical specimens in same shipment, please put RNA extracts and clinical specimens in separate biohazard bags.</li> <li>Maintain proper infection control when handling specimens. Guidance can be found at <a href="https://www.cdc.gov/infectioncontrol/guidelines/isolation/index.html#a4">https://www.cdc.gov/infectioncontrol/guidelines/isolation/index.html#a4</a>.</li> <li>If shipping specimens through a courier, specimens may be shipped Monday-Thursday for receipt at the DSHS Laboratory Tuesday-Friday. Weekend delivery will be considered on a case by case basis and requires pre-approval by DSHS. Contact EAIDU epidemiologists for pre-approval.</li> <li>Place the specimen container in a zip-lock style biohazard bag and securely seal the bag. Place a hardcopy of the line list in the outer sleeve of the bag or attach it to the outside of the bag (Note: DO NOT seal the line list inside the biohazard bag with the specimen as we cannot process potentially contaminated forms). Put the bag or bags in a Styrofoam insulated cardboard box for shipping.</li> <li>Follow international Air Transportation Regulations for shipping of Category "B" Biological Substances - Infectious Subs. brochure - PHMSA.</li> <li>Ship to the physical address: TX DSHS Laboratory Services, ATTN: Walter Douglass 512-776-7569, 1100 W. 49th Street, Austin TX, 78756.</li> <li< th=""></li<></ul>
Results Reporting	<ul> <li>If the submitter/provider requests the results, the results will be added to the submitted line list and emailed back to the submitter by the DSHS Laboratory.</li> </ul>